
Lung Regeneration And Organoids

Abstract

In recent times there has been a great increase in the number of chronic lung diseases and lungs being one of the vital soft tissue organs of the body need transplantation or excision of the damaged tissue^{1,2}. Though recent medical advances and transplantation procedures have saved numerous lives their efficiency to provide a better life outcome are suboptimal. Stem cells, induced pluripotent stem cells and tissue regenerative scaffolds have made huge strides to improve patient therapies for the same^{2–5} but in case of lungs a 3D tissue model mimicking the functioning of an actual organ is absolutely necessary because in disease modelling, developmental studies and regenerative medicine^{4,6–8}, just cells cultures are not enough to understand the diseases in-vivo or regrow a fully functioning 3D tissue or organ^{1,8–10}. This where organoids come into the picture, they are stem cell-derived 3D structures that are supported by an extracellular matrix and contain multiple cell types whose spatial arrangement and interactions mimic those of a native organ.⁷ Aim of this article is to explore the different and novel strategies^{3,4,9,11–13} used for lung regeneration and their use or interplay with lung organoids, hence ultimately determining their impact in regenerative medicine.

Introduction: -

Anatomy of human lung and lung diseases:

A brief outline on the structure and physiological function of the human lung. Focus on pointing of the most vulnerable and important parts or areas of the lung tissue¹.

And leading to impairment of these parts either due to physiological deterioration, invasion of pathogens etc. leads to various chronic diseases like Pneumonia, lung collapse, COPD, lung cancer etc^{2–4}.

Lung organoids

They are stem cell-derived 3D structures that are supported by an extracellular matrix and contain multiple cell types whose spatial arrangement and interactions mimic those of a native organ. Highlight on the different cellular population necessary for its proper formation^{4–9}

Purpose is to define and understand what organoids are and their importance so as to be used in regenerative medicine.

Regeneration of lungs

Principles governing the natural process of regeneration in lungs and tissue turn-over. Tissue turn-over is obviously quite low in human adult lungs. There are complex signaling pathways working and communicating between many different cell types like facultative lung progenitor cells, basal cells, lung progenitor and stem cells, allowing them to become activated, get

recruited at the site of injury and ultimately repair the tissue^{1,3,10,11}.

Current Therapeutics in Lung Regeneration and Repair

Medical strategies in lung repair

Many people suffer from chronic lung diseases where the possibility to heal or repair the damaged tissue is next to impossible job, hence in today's medicinal world doctors use procedures like lung grafting – to partially or completely replace the damaged tissue by a healthy one obtained from a donor^{12–14}.

Therapeutic strategies in lung regeneration

Stem cell mediated therapies have long been looked up as the alternative for lung regeneration and repair^{8,10,11,15}.

Problems like immune reactions to grafts, organ failures, unaccounted or uncontrollable cell growth and many more are often faced while implementing current therapeutic strategies^{8,11–13,16}

Culture Systems for Organoids Development

Cell cultures where the cells are grown on a monolayer, either to observe their behavior or obtain a specific cell type lineage through directing cell specific differentiation and proliferation by providing different factors^{7,8,17–19}

The air-liquid interface culture helps us derive a more lung like environment and drives airway epithelial cells to proliferate and differentiate in-vitro^{2,7,20,21}.

New approaches allowing to attain more organ like in-vivo environment thereby helping to obtain organoids. Cells are grown in a three dimensional either natural or synthetically fabricated scaffold where the cells differentiation and proliferation is tightly regulated by the extrinsic factors^{5,7,8,21,22}.

Different Approaches to form Lung Organoids

Lung organoids have known to be obtained from many different sources of cells, the most prominent and successful ones are discussed and enlisted below. Though trying to achieve the same result of a functional organoid, they do differ and, in the process, how they are obtained

3D ALI organoids obtained from stem or different progenitor population were retaining their regenerative capabilities both in-vivo and in-vitro thereby providing great solutions as tissue/scaffold replacement so as to repair or regenerate lung tissue^{2,8,24,35–37}

To treat human diseases, one must have a thorough understanding of the same and these studies give much more accurate results when they are carried in systems which mimic the actual tissue or organ as closely as possible, like an organoid. Thereby also reducing animal and human sacrifices too^{4,7,8,31,36,38}:

Post understanding the ailment it is necessary to test and develop a cure for it, most of times in the form of drugs. That's why organoid systems allow to assess the real time and immediate effects of the drug in that specific tissue, seeing their effect on multiple cell lineages and also on the crosstalk between them^{8,16,39–45}

Outlook

The new approach with stem and progenitor cells and their use in 3D cultures have enabled to researchers to produce reproducible and regenerative organoid systems which closely mimic the tissue of desire. Opening up endless possibilities and unique solutions to many of our recent problems.

References

1. Crapo, J. D., Barry, B. E., Gehr, P., Bachofen, M. & Weibel, E. R. Cell number and cell characteristics of the normal human lung. *Am. Rev. Respir. Dis.* 126, 332–337 (1982).
2. Schilders, K. A. A. et al. Regeneration of the lung: Lung stem cells and the development of lung mimicking devices. *Respir. Res.* 17, 1–16 (2016).
3. Lee, J. H. & Rawlins, E. L. Developmental mechanisms and adult stem cells for therapeutic lung regeneration. *Dev. Biol.* 433, 166–176 (2018).
4. Nadkarni, R. R., Abed, S. & Draper, J. S. Organoids as a model system for studying human lung development and disease. *Biochem. Biophys. Res. Commun.* 473, 675–682 (2016).
5. Tan, Q., Choi, K. M., Sicard, D. & Tschumperlin, D. J. Human airway organoid engineering as a step toward lung regeneration and disease modeling. *Biomaterials* 113, 118–132 (2017).
6. Miller, A. J. et al. Generation of lung organoids from human pluripotent stem cells in vitro. *Nat. Protoc.* 14, 518–540 (2019).
7. Choi, J., Ilich, E. & Lee, J. H. Organogenesis of adult lung in a dish: Differentiation, disease and therapy. *Dev. Biol.* 420, 278–286 (2016).
8. Barkauskas, C. E. et al. Lung organoids: Current uses and future promise. *Dev.* 144, 986–997 (2017).
9. Fatehullah, A., Tan, S. H. & Barker, N. Organoids as an in vitro model of human development and disease. *Nat. Cell Biol.* 18, 246–254 (2016).
10. Akram, K. M., Patel, N., Spiteri, M. A. & Forsyth, N. R. Lung regeneration: Endogenous and exogenous stem cell mediated therapeutic approaches. *Int. J. Mol. Sci.* 17, (2016).
11. Chambers, R. C. Abnormal wound healing responses in pulmonary fibrosis: Focus on coagulation signalling. *Eur. Respir. Rev.* 17, 130–137 (2008).
12. Afonso Júnior, J. E. et al. Lung transplantation *Transplante pulmonar*. *Einstein* 13, 297–304 (2015).
13. Gadre, S., Turowski, J. & Budev, M. Overview of Lung Transplantation, Heart-Lung Transplantation, Liver-Lung Transplantation, and Combined Hematopoietic Stem Cell Transplantation and Lung Transplantation. *Clin. Chest Med.* 38, 623–640 (2017).
14. Resection, S. & Cancer, I. I. L. Wedge Resection Versus Anatomic Resection : Extent of. 426–433 (2017).
15. Chen, F. & Fine, A. Stem Cells in Lung Injury and Repair. *Am. J. Pathol.* 186, 2544–2550 (2016).
16. Newman, S. P. Drug delivery to the lungs: Challenges and opportunities. *Ther. Deliv.* 8,

647–661 (2017).

17. Duval, K. et al. Modeling physiological events in 2D vs. 3D cell culture. *Physiology* 32, 266–277 (2017).
18. Htwe, S. S. et al. Investigating NF- κ B signaling in lung fibroblasts in 2D and 3D culture systems. *Respir. Res.* 16, 1–9 (2015).
19. Breslin, S. & O'Driscoll, L. The relevance of using 3D cell cultures, in addition to 2D monolayer cultures, when evaluating breast cancer drug sensitivity and resistance. *Oncotarget* 7, 45745–45756 (2016).
20. Hiemstra, P. S., Tetley, T. D. & Janes, S. M. Airway and alveolar epithelial cells in culture. *Eur. Respir. J.* 1900742 (2019). doi:10.1183/13993003.00742-2019
21. Chen, S. & Schoen, J. liquid interface cell culture : From airway epithelium to the female reproductive tract. 54, 38–45 (2019).
22. Jain, R. et al. Plasticity of Hopx+ type I alveolar cells to regenerate type II cells in the lung. *Nat. Commun.* 6, (2015).
23. Dye, B. R. et al. A bioengineered niche promotes in vivo engraftment and maturation of pluripotent stem cell derived human lung organoids. *Elife* 5, 1–18 (2016).
24. Gkatzis, K., Taghizadeh, S., Huh, D., Stainier, D. Y. R. & Bellusci, S. Use of three-dimensional organoids and lung-on-a-chip methods to study lung development, regeneration and disease. *Eur. Respir. J.* 52, (2018).
25. Ghosh, M. et al. Human tracheobronchial basal cells: Normal versus remodeling/repairing phenotypes in vivo and in vitro. *Am. J. Respir. Cell Mol. Biol.* 49, 1127–1134 (2013).
26. Zacharias, W. J. et al. Regeneration of the lung alveolus by an evolutionarily conserved epithelial progenitor. *Nature* 555, 251–255 (2018).
27. Ng-Blichfeldt, J. P. et al. Retinoic acid signaling balances adult distal lung epithelial progenitor cell growth and differentiation. *EBioMedicine* 36, 461–474 (2018).
28. Hynds, R. E., Bonfanti, P. & Janes, S. M. Regenerating human epithelia with cultured stem cells: feeder cells, organoids and beyond. *EMBO Mol. Med.* 10, 139–150 (2018).
29. Liu, X., Driskell, R. R. & Engelhardt, J. F. Airway Glandular Development and Stem Cells. *Curr. Top. Dev. Biol.* 64, 33–56 (2004).
30. Usselman, C. W. N. S. S. J. R. B. ?????? HHS Public Access. *Physiol. Behav.* 176, 139–148 (2017).
31. Strikoudis, A. et al. Modeling of Fibrotic Lung Disease Using 3D Organoids Derived from Human Pluripotent Stem Cells. *Cell Rep.* 27, 3709-3723.e5 (2019).
32. Zimmermann, B. Lung organoid culture. *Differentiation* 36, 86–109 (1987).
33. Yamamoto, Y. et al. Long-term expansion of alveolar stem cells derived from human iPS cells in organoids. *Nat. Methods* 14, 1097–1106 (2017).
34. Snyder, J. M., Johnston, J. M. & Mendelson, C. R. Differentiation of type II cells of human fetal lung in vitro. *Cell Tissue Res.* 220, 17–25 (1981).
35. Clevers, H. Modeling Development and Disease with Organoids. *Cell* 165, 1586–1597 (2016).
36. Mathis, C. et al. Human bronchial epithelial cells exposed in vitro to cigarette smoke at the air-liquid interface resemble bronchial epithelium from human smokers. *Am. J. Physiol. - Lung Cell. Mol. Physiol.* 304, (2013).
37. Diao, J. et al. Sweat gland organoids contribute to cutaneous wound healing and sweat gland regeneration. *Cell Death Dis.* 10, (2019).
38. Skronska-Wasek, W. et al. Reduced frizzled receptor 4 expression prevents WNT/ β -catenin-driven alveolar lung repair in chronic obstructive pulmonary disease. *Am. J. Respir. Crit. Care Med.* 196, 172–185 (2017).

-
39. Paolicelli, G. et al. Using lung organoids to investigation epithelial barrier complexity and IL-7 signaling during respiratory infection. *Front. Immunol.* 10, 1–6 (2019).
 40. Fang, Y. & Eglen, R. M. Three-Dimensional Cell Cultures in Drug Discovery and Development. *SLAS Discov.* 22, 456–472 (2017).
 41. Jung, D. J. et al. A one-stop microfluidic-based lung cancer organoid culture platform for testing drug sensitivity. *Lab Chip* 19, 2854–2865 (2019).
 42. Xu, H. et al. Organoid technology in disease modelling, drug development, personalized treatment and regeneration medicine. *Exp. Hematol. Oncol.* 7, 1–12 (2018).
 43. Weeber, F., Ooft, S. N., Dijkstra, K. K. & Voest, E. E. Tumor Organoids as a Pre-clinical Cancer Model for Drug Discovery. *Cell Chem. Biol.* 24, 1092–1100 (2017).
 44. Kim, M. et al. Patient-derived lung cancer organoids as in vitro cancer models for therapeutic screening. *Nat. Commun.* 10, (2019).
 45. Xinaris, C. Organoids for replacement therapy. *Curr. Opin. Organ Transplant.* 24, 555–561 (2019).

edubirdie.com